



Metagenome analysis of low-biomass samples: A JGI User Facility Perspective

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Metagenome Program Head

NASA Planetary Protection Workshop
November 21, 2024



The DOE Joint Genome Institute at a glance

Integrative Genomics Building (IGB)



JGI MISSION:

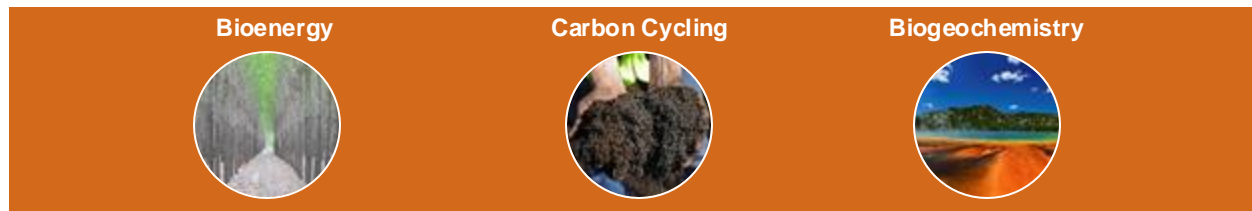
Provide advanced genomic capabilities, large-scale data, and professional expertise to support the global research community in studies of complex biological and environmental systems. We optimize our service to the community through responsibly managing our people and resources.

A U.S. Department of Energy Office of Science User Facility

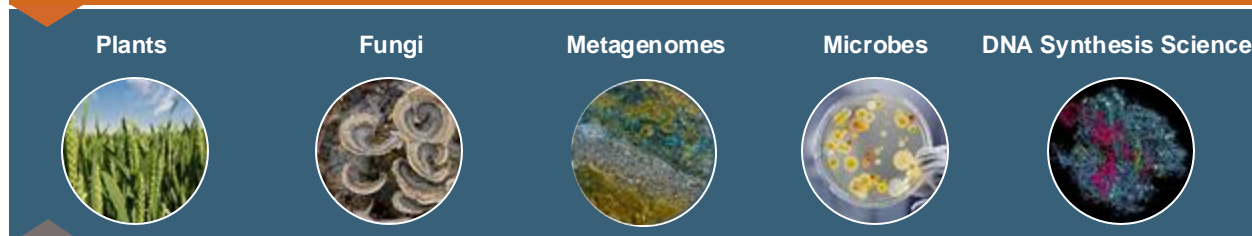
- JGI established in 1997, User facility from 2004
- IGB in Berkeley, CA opened in Dec 2019
- ~260 staff; ~\$92M annual funding

Mission Areas, Programs & Infrastructure


Mission Areas



Programs



Infrastructure

 <p>DNA Sequencing</p> <ul style="list-style-type: none"> • Illumina platforms: NovaSeq, HiSeq, MiSeq, Next Seq • PacBio platforms: RSII, Sequel • Oxford Nanopore platforms: MinION, PromethION 	 <p>Advanced Genomic Technologies</p> <ul style="list-style-type: none"> • Single-cell Seq • Metabolomics • ChIP-Seq, Tn-Seq, DAP-Seq • Epigenomics • Transcriptomics 	 <p>Computational Analysis</p> <ul style="list-style-type: none"> • Assembly/Annotation/Analysis • High-Performance Computing • Big Data Integration & Analysis Systems 	 <p>DNA Synthesis</p> <ul style="list-style-type: none"> • Design & Pathway Assembly • Host Engineering
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JGI Metagenome Program - Products

Metagenome

shotgun DNA sequencing
Phylogenetic diversity

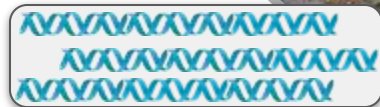
Minimal draft
(~35M reads)



Standard draft
(~150M reads)



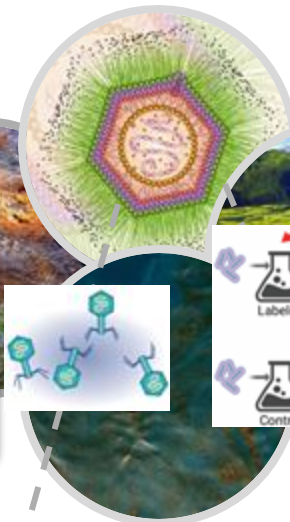
Improved draft
(PacBio Sequel II)



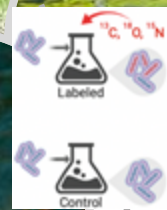
Viral metagenome
(ss & dsDNA)



Function driven
metagenomics



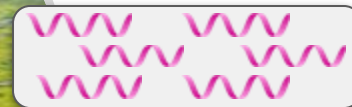
Viral-host
interactions



Metatranscriptome

shotgun RNA sequencing
RNA-Seq

Ribo-depletion
or
PolyA selection



microbial ecology

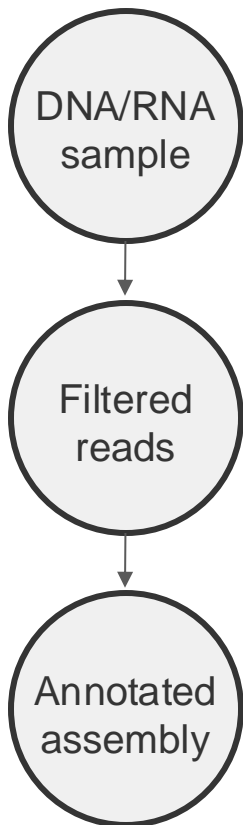


SIP metagenomes
(density fractions
enrichment
analysis)

**CSP Annual Call: 10Tb cap
(up to 50Tb for multi-PI projects)**

**FICUS / CSP New Investigator:
3Tb cap**

JGI Metagenome Program – Standard output



- Sample QC
- Library preparation
- Sequencing & QC

- Assembly (metaSPAdes for metagenomes)
- Read mapping
- Genome binning (Metabat2) and QC
- Functional annotation (IMG pipeline)

- Data release through JGI Data Portal (download) and IMG (exploration/analysis) plus IMG/VR (Virus genomes)



DOE JGI Metagenome Workflow

Authors: Alicia Clum, Marcel Huntemann, Brian Bushnell, Brian Foster, Bryce Foster, Simon Roux, Patrick P. Hajek, Neha Varghese, Supratim Mukherjee, T. B. K. Reddy, Chris Daum, Yuko Yoshinaga, Ronan O'Malley, Rekha Seshadri, Nikos C. Kyrpides, Emiley A. Elie-Fadrosch, I-Min A. Chen, Alex Copeland, Natalia N. Ivanova



Data Portal



IMG/M

INTEGRATED MICROBIAL GENOMES & MICROBIOMES



IMG/VR

INTEGRATED MICROBIAL GENOMES / VIRUS

Library prep for metagenomes

Sample Registration



Sample QC: volume concentration quality



Two paths for library creation



384-well automation: fixed PCR cycles

- Illumina PCR-free WGS libraries
- Illumina low input (1 ng) WGS libraries (9 cycles of PCR)
- Illumina RNA-seq (1 ug input 8 cycles of PCR, 100 ng input with 14 cycles of PCR)

8-striptide manual libraries:

Ultra-low input (undetectable to <1 ng) - flexible PCR cycles (>15) &:

- Illumina ss/dsDNA-seq (1 ng) with flexible PCR cycles (>12)
- PacBio PCR-free long reads WGS (1 ug)
- PacBio low input (20 ng) long reads WGS with flexible PCR cycles (>12)



Example of ultra-low input samples: ice core viruses

- **Viromes from ice-cores**

- Low input, contamination-prone
- First view of viruses “trapped in ice”

- **High level of genetic novelty**

- Novel phages linked to dominant ice-adapted bacteria
- Remote similarity to plant-associated, soil, freshwater sequences

User Programs

CSP# 503428



Zhiping Zhong



Matthew Sullivan

Smithsonian
MAGAZINE

SMARTNEWS Keeping you current

Scientists Extract 15,000-Year-Old
Viruses From Tibetan Glacier



NEWS

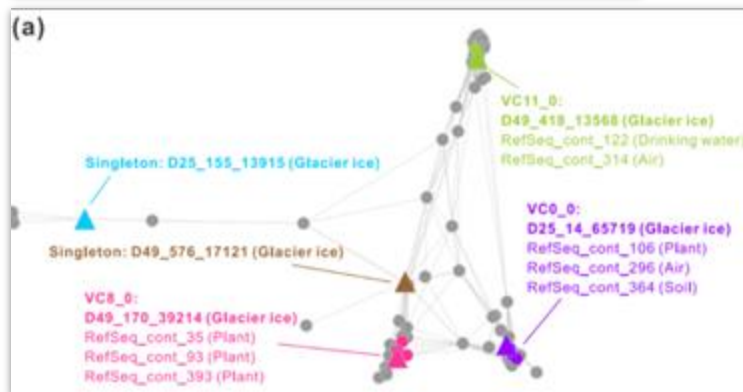
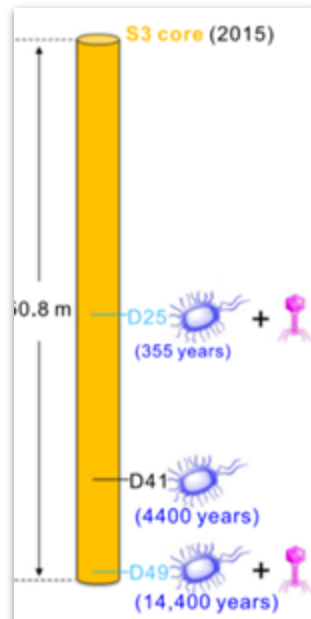
15,000-year-old viruses never seen by humans
discovered in glacier ice

WORLD

Scientists discover more than 30
viruses frozen in ice, most never
seen before

Jordan Mendoza USA TODAY

Published 2:49 p.m. ET Jul. 21, 2021 | Updated 6:01 p.m. ET Jul. 21, 2021



Primary template-directed amplification: addressing coverage bias



Charles Gawad



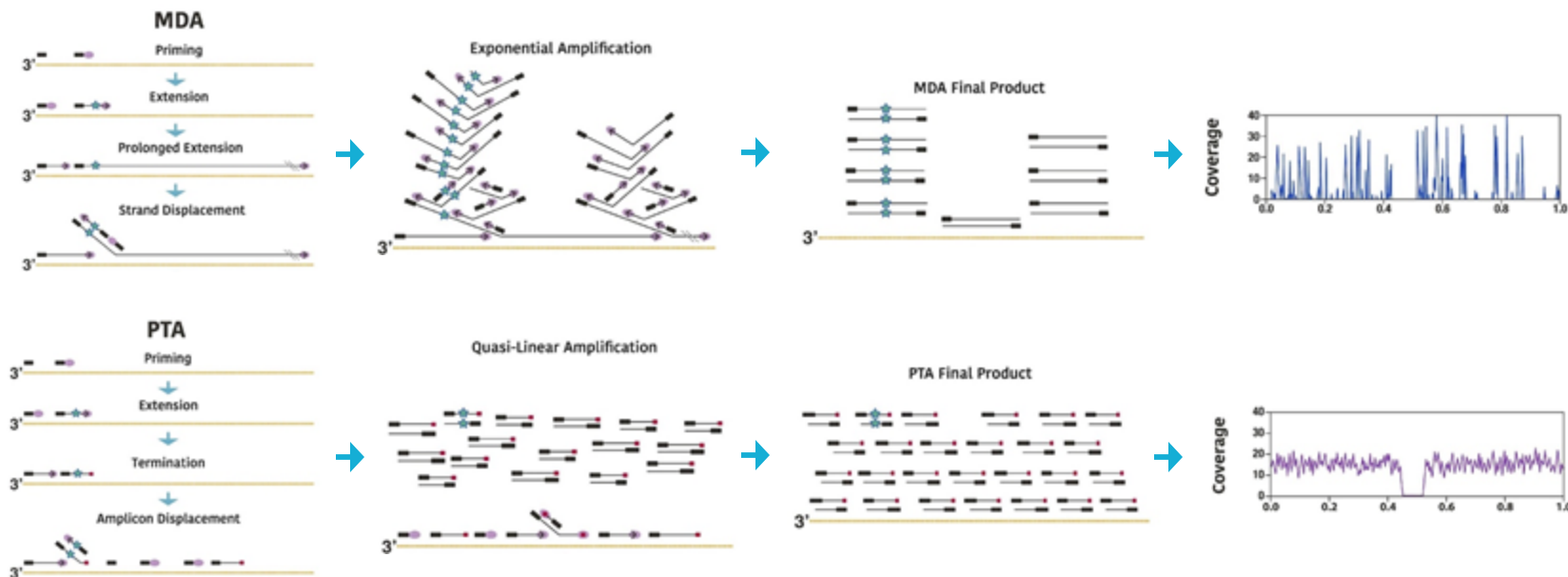
Veronica Gonzalez-Pena



Comparison of PTA to MDA on human cells

Addition of terminators limits over-amplification

Quasi-linear amplification improves coverage uniformity



Can PTA help improve the quality of microbial single cells?

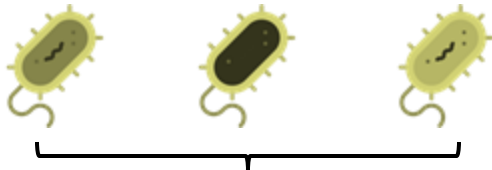


Microbial isolates
(n = 34)

E. coli
(n = 12)

P. putida
(n = 12)

B. subtilis
(n = 10)



PTA

MDA

WGA-X

Environmental microbes
(n = 185)



Mountain View Sough
(n = 97)



LBNL soil
(n = 27)

PTA

MDA

WGA-X



ISME Communications, 2024, 4(1), ycae085
<https://doi.org/10.1093/ismeco/ycae085>
Advance access publication: 12 July 2024
Brief Communication

scMicrobe PTA: near complete genomes from single bacterial cells

Robert M. Bowers^{1,2}, Veronica Gonzalez-Pena^{2,3}, Kartika Wardhani², Danielle Goudeau¹, Matthew James Blow¹, Daniel Udway¹, David Klein², Albert C. Vill³, Ilana L. Brito³, Tanja Woyke¹, Rex R. Malmstrom^{2,3,4,5}, Charles Gawad^{2,3,4,5}



Bob
Bowers



Danielle
Goudeau



Tanja
Woyke



Rex
Malmstrom

Can PTA help improve the quality of microbial single cells?

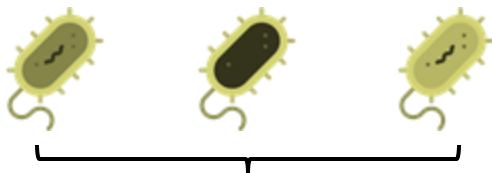


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E. coli
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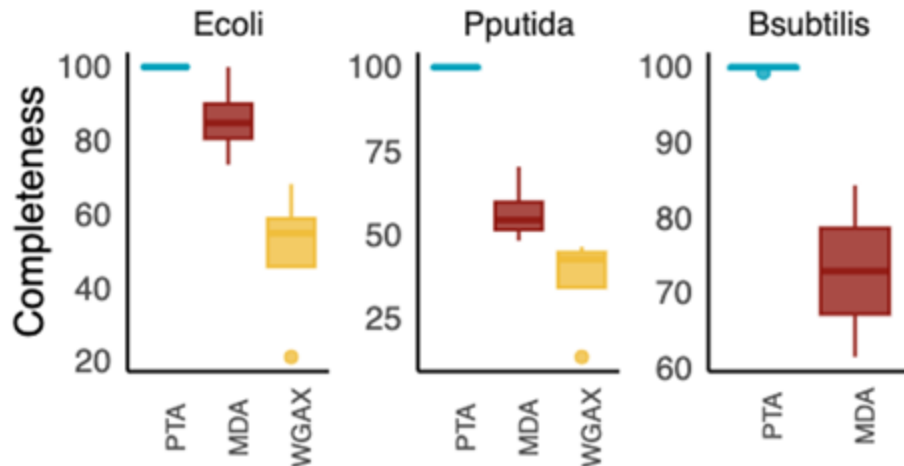
PTA

MDA

WGA-X

Analyses

- Genome coverage uniformity
- Genome quality statistics



Environmental benchmarking: Overall quality improves

Environmental microbes (n = 185)



Mountain View Sough
(n = 97)



LBNL soil
(n = 27)

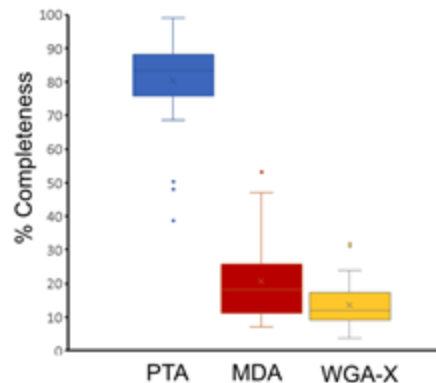
PTA

MDA

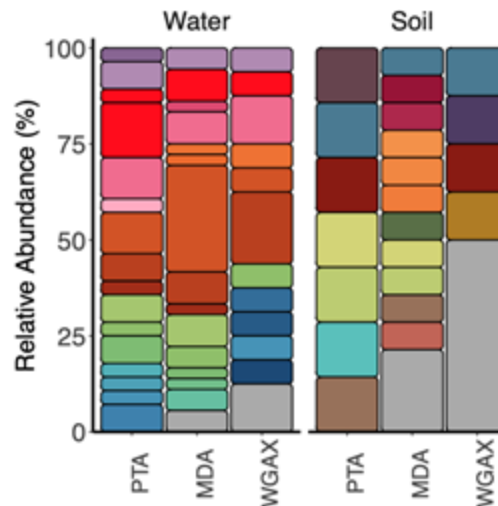
WGA-X

Analyses

- Genome quality statistics
- Taxonomic composition



- Significant improvement in SAG completeness
- Moderate contamination in PTA'd SAGs
- Increase in medium- and high-quality SAGs



Taxonomy

- | | |
|-------------------------|-------------------------|
| Ac; Pyrinomonadaceae | Bd; Bdellovibrionaceae |
| Ac; UBA2999 | Fi; Streptococcaceae |
| Ac; UBA11606 | Ga; Methylophilaceae |
| Ac; Microbacteriaceae | Ga; SG8-39 |
| Ac; Gaiellaceae | Ga; UKL13-2 |
| Al; Dongiaceae | Ga; Sedimenticolaceae |
| Al; HIMB59 | Ga; Alteromonadaceae |
| Al; Pelagibacteraceae | Ga; Nevskiaceae |
| Al; UBA1172 | Ga; Thioglobaceae |
| Al; Rhodobacteraceae | Ga; unknown |
| Al; unknown | Ga; DSM-100316 |
| Al; Sphingomonadaceae | Ga; Halieaceae |
| Ba; B-17BO | Ga; HTCC2089 |
| Ba; Cyclobacteriaceae | Ga; Litoricolaceae |
| Ba; CAIKAF01 | Ga; Nitrincolaceae |
| Ba; Flavobacteriaceae | Ga; Pseudohongiellaceae |
| Ba; Schleiferiaceae | Ga; UBA4575 |
| Ba; unknown | Ge; Gemmatimonadaceae |
| Ba; UKL13-3 | Ve; Opitutaceae |
| Ba; Sphingobacteriaceae | unassigned |
| Ba; Balneolaceae | |